

MASSONS TRICHROM STAIN KIT (SS010-25)

INTENDED USE

BioMarq Masson's Trichrome Stain Kit is used for *in vitro* diagnostic use. It is a qualitative histologic stain used to study connective tissue, muscle and collagen fibers in formalin-fixed, paraffin-embedded tissue. Masson's Trichrome stain is used to study muscular pathologies (muscular dystrophy), cardiac pathologies (infarct), hepatic pathologies (cirrhosis) or kidney pathologies (glomerular fibrosis). It can also be used to detect and analyze routinely tumors on hepatic and kidney biopsies. The results using this product should be interpreted by a qualified pathologist in conjunction with the patient's relevant clinical history, other diagnostic tests and proper controls.

PRODUCT DESCRIPTION

Masson's trichrome staining involves tricolor staining, where in nuclei stained with Hematoxylin and are stable for decolorizing agents. Further on Biebrich scarlet-acid fuchsin solution stain the tissue components red. The tissue components which are less permeable for acids (collagen) get de-stained with the treatment acid (phosphomolybdic/phosphotungstic acid) finally the de-stained collagen fibers are re-stained with aniline blue.

KIT CONTENTS

Bouin's Solution (SS026-8D)	1X8ml
Weigert's Hematoxylin A (SS031-5D)	1X5ml
Weigert's Hematoxylin B (SS032-5D)	1X5ml
Biebrich Scarlet-Acid Fuchsin (SS033-8D)	1X8ml
Acetic Acid 10% (SS055-8D)	1X8ml
Phospho molybdo tungstic Acid (SS052-8D)	1X8ml
Aniline Blue (SS035-8D)	1X8ml

MATERIALS REQUIRED BUT NOT PROVIDED

Microscope slides
 Xylene/ Xylene substitute
 Reagent alcohol/Ethanol
 Deionized or distilled water
 Wash buffer
 Tissue controls
 Mounting Solutions

STORAGE AND STABILITY

Store at 2-8°C. Do not freeze. Not to be used beyond the expiration date prescribed on label.

SPECIMEN PREPARATION

Fixation: Bouin's is preferred, 10% formalin.
Paraffin Sections: Cut sections at 4 microns.

RECOMMENDED PROTOCOL

Deparaffinize with xylene/ xylene substitute and rehydrate through graded alcohols to deionized water. Wash the slide with SS Wash Solution for 3 times.

Fix the tissue with 500ul Bouin's solution at 40° C for 30 min to improve staining quality. If required apply cover slip. Wash the slides thoroughly with SS wash solution till the yellow color is removed.

Stain the slides 300ul of Weigert's Working Solution for 10 min. Wash the slides with SS Wash Solution for 3 times.

Stain the slides 300ul of Biebrich Scarlet solution for 10 -15 min. Wash the slides with SS Wash Solution for 3 times.

Differentiate the slides in 300ul of Phospho molybdo-tungstic Acid solution for 10 -15 min.

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Blot the slides and immediately stain the slides with 300ul of aniline blue solution for 3-5min. Wash the slides with SS Wash Solution for 3 times.

Differentiate the slides with 1% acetic acid for 3 min. Wash the slides with SS Wash Solution for 3 times.

Dehydrate, clear in xylene or its substitute & mount the slides with BioMarq XY-Mount (Catalog No MS002) or using BioMarq T-Mount (Catalog No MS003).

TECHNICAL NOTE

Bring all reagents to RT before use

Gently invert all reagents prior to use

Thickness of section may affect the intensity of staining

Follow the protocol recommendations provided in the data sheet. If atypical results occur, contact BioMarq Technical Support at 040-29702960.

RECOMMENDED CONTROL

COLON, STOMACH, INTESTINE AND SKIN

Users can also procure the Qualified Positive Control Slides available from BioMarq for their Quality Control purpose.

STAINING INTERPRETATION

Muscle, cytoplasm and keratin - Red
Nuclei - purple-Violet and
Collagen fibers – Blue.

For image please visit our web site www.biomarq.net

PRECAUTIONS

Specimens should be handled carefully before and after the assay to avoid transmission of infection and disposed of with proper precautions

When working with Special Stain chemicals, always wear a suitable lab coat, disposable gloves, and protective goggles

For detailed safety information related to BioMarq Products, please refer to appropriate safety data sheets (SDS) available online at www.biomarq.net

REFERENCES

1. Calvi EN et al, An experimental model for the study of collagen fibers in skeletal muscle, Acta Cir Bras. 2012 Oct.
2. Ayen, E et al, Histological and histomorphological changes of different regions of oviduct during follicular and luteal phase of estrus cycle in adult, Iranian J. Vet. Res, 2012.
3. Meinen et al, Linker molecules between laminins and dystroglycan ameliorate laminin-alpha2-deficient muscular dystrophy at all disease stages. The Journal of cell Biology, 2007.
4. Hena SA et al, Demonstration of Muscle Fibre Types Using Masson Trichrome Stain from Deltoideus Muscles of One-humped Camel (Camelus Dromedarius), J Mol Histol Med Physiol, 2017 January 20.